

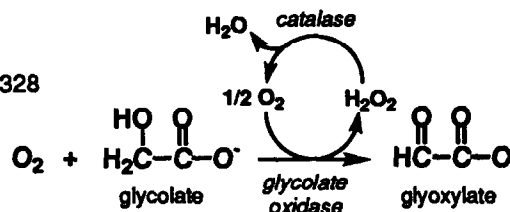
GRAPHICAL ABSTRACTS

BioMed. Chem. 1994, 2, 371

Glyoxylic Acid Production Using Immobilized Glycolate Oxidase and Catalase

John E. Seip, Susan K. Fager, John E. Gavagan,
David L. Anton, and Robert Di Cosimo*
CR&D Dept., E. I. du Pont de Nemours & Co.,
Experimental Station, PO Box 80328, Wilmington, DE 19880-0328

A variety of enzyme immobilization methods have been examined for the preparation of a catalyst for the oxidation of glycolic acid to glyoxylic acid in 99 % yield.

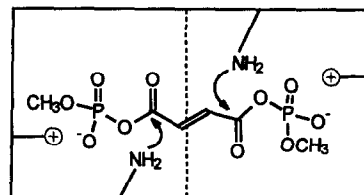


BioMed. Chem. 1994, 2, 379

INACTIVATION OF D-3-HYDROXYBUTYRATE DEHYDROGENASE BY FUMAROYL BIS(METHYL PHOSPHATE)

Ronald Kluger* and Stephen Bearne, *Department of Chemistry, University of Toronto, Toronto, Canada M5S 1A1*

Abstract: Fumaroyl bis(methyl phosphate) inactivates D-3-hydroxybutyrate dehydrogenase while cross-linking the two subunits of the enzyme.



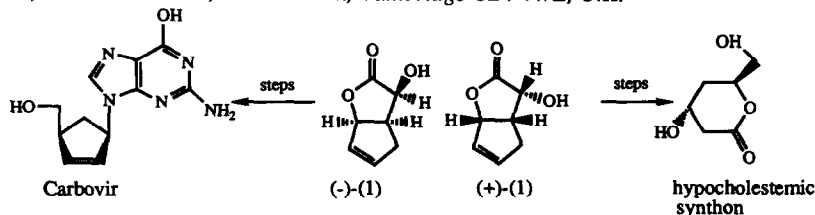
Enzyme-Catalysed Kinetic Resolution of 4-endo-Hydroxy-2-oxabicyclo[3.3.0]-oct-7-ene-3-one and Employment of the

Pure Enantiomers for the Synthesis of Anti-viral and Hypocholesteremic Agents.

R. A. MacKeith, R. McCague, H. F. Olivo, S. M. Roberts, S. J. C. Taylor and H. Xiong

^aDepartment of Chemistry, Exeter University, Exeter, Devon EX4 4QD, U.K.

^bResearch Division, Chiroscience Ltd, Science Park, Cambridge CB4 4WE, U.K.



BioMed. Chem. 1994, 2, 387

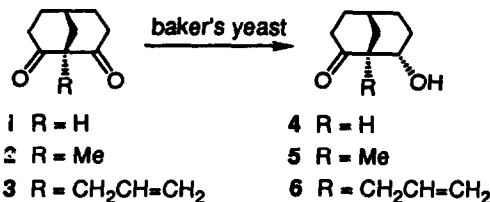
Reduction of Bicyclo[3.3.1]nonane-2,8-diones with Baker's Yeast

Kenji Mori,*^a Shuichi Takayama^a and Masaru Kido^b

^aDepartment of Agricultural Chemistry, The University of Tokyo, Yayoi 1-1-1, Bunkyo-ku, Tokyo 113, Japan

^bAnalytical Chemistry Laboratory, 2nd Tokushima Institute of New Drug Research, Otsuka Pharmaceutical Co., Ltd., Kawauchi, Tokushima 771-01, Japan

Reduction of bicyclo[3.3.1]nonane-2,8-dione(1) and its homologues 2 and 3 with baker's yeast affords (1R, 5S, 8S)-8-hydroxybicyclo[3.3.1]nonan-2-one (4) and its homologues 5 and 6.

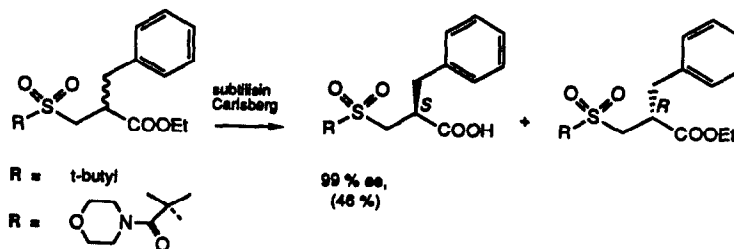


BioMed. Chem. 1994, 2, 395

Large Scale Preparation of Chiral Building Blocks for P₃ Site of Renin Inhibitors

BioMed. Chem. 1994, 2, 403

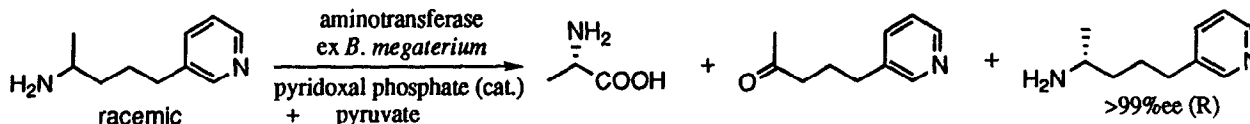
S. Doswald, H. Esterman, E. Kupfer, H. Stadler, W. Walther, T. Weisbrod, B. Wirz*, W. Wostl
Pharma Research, New Technologies, ^aDept. of Microbiology or ^dPhysics; Pharma Operations, Manufacturing, ^bChemical Production or ^cProcess Development; ^ePharma Research, Cardiovascular Diseases; F. Hoffman-La Roche Ltd, 4002 Basel, Switzerland



AMINOTRANSFERASE CATALYSIS APPLIED TO THE SYNTHESIS OF A PAF ANTAGONIST

BioMed. Chem. 1994, 2, 411

David L. Coffen, Masami Okabe, Ruen Chu Sun, Seju Lee and George W. J. Matcham, Roche Research Center, Hoffmann-La Roche Inc., Nutley, NJ 07110 Celgene Corporation, Warren, NJ 07059

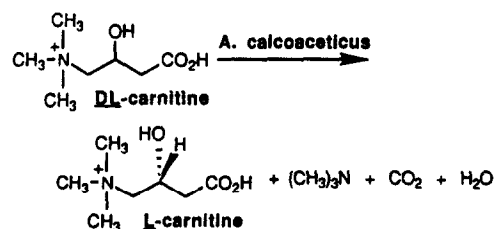


L-CARNITINE VIA ENZYME-CATALYZED OXIDATIVE KINETIC RESOLUTION.

BioMed. Chem. 1994, 2, 415

Dennis Ditullio, David Anderson, Ching-Shih Chen, and Charles J. Sih*
School of Pharmacy, University of Wisconsin, Madison, WI 53706 U.S.A.

L-Carnitine of high optical purity was prepared via kinetic resolution using a mutant strain of *Acinetobacter calcoaceticus* ATCC 39647. This organism preferentially metabolized the D-enantiomer of the racemate to furnish L-carnitine. Recovery of L-carnitine was 93%, providing a total weight yield of 46.5% in 92% enantiomeric excess. The mode of degradation of carnitine was shown to proceed via a monooxygenase-catalyzed oxidative cleavage resulting in the formation of trimethylamine and malic acid.

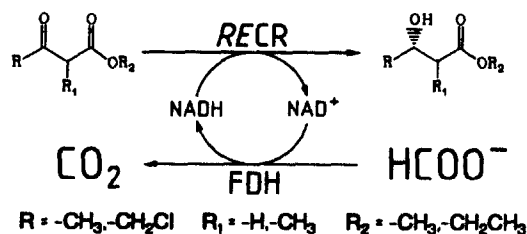


A KINETIC STUDY AND APPLICATION OF A NOVEL CARBONYL REDUCTASE ISOLATED FROM RHODOCOCCUS ERYTHROPOLIS

BioMed. Chem. 1994, 2, 421

Thomas Zelinski and Maria-Regina Kula
Institut für Enzymtechnologie der
Heinrich-Heine-Universität Düsseldorf,
KFA Jülich, PO Box 2050, 52404 Jülich, FRG

Summary: A NADH-dependent Carbonyl Reductase which reduce a broad range of carbonyl compounds to (S)-hydroxy compounds. Application of the enzyme in preparative synthesis of 3-hydroxyacid esters.

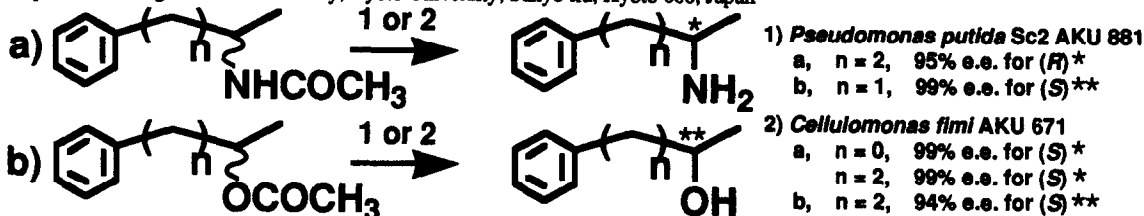


Enzymatic asymmetric synthesis of α -methyl arylalkylamines and α -methyl arylalkylalcohols by arylalkyl acylamidases.

Jun Ogawa, Sakayu Shimizu and Hideaki Yamada,

Department of Agricultural Chemistry, Kyoto University, Sakyo-ku, Kyoto 606, Japan

BioMed. Chem. 1994, 2, 429

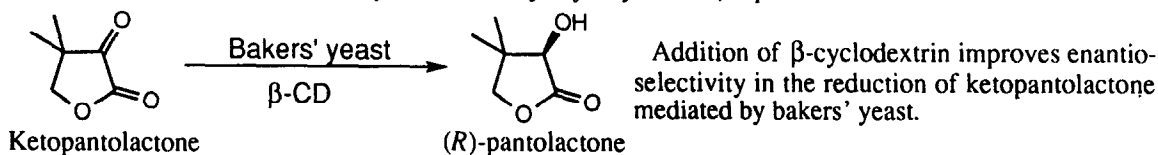


EFFECT OF CYCLODEXTRIN ON IMPROVEMENT OF ENANTIOSELECTIVITY IN THE REDUCTION OF KETOPANTOLACTONE WITH BAKERS' YEAST

Kaoru Nakamura, Shin-ichi Kondo, Atsuyoshi Ohno*

Institute for Chemical Research, Kyoto University, Uji, Kyoto 611, Japan

BioMed. Chem. 1994, 2, 433



CHEMICAL AND ENZYME-CATALYSED SYNTHESIS OF QUINOLINE HYDRATES.

R. Agarwal, D. R. Boyd, N. D. Sharma, R. A. S. McMordie, and H. P. Porter,
School of Chemistry, The Queen's University of Belfast, Belfast BT9 5AG, UK.

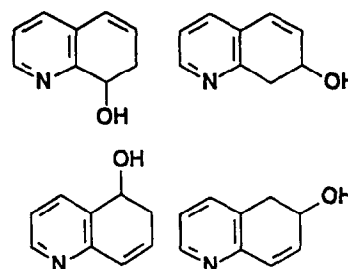
B. van Ommen, and P. J. van Bladeren,

Department of Biological Toxicology, TNO Toxicology and Nutrition

Institute, P.O. Box 360, 3700 AJ Zeist, The Netherlands.

BioMed. Chem. 1994, 2, 439

Four arene hydrates of quinoline have been synthesised by chemical and enzyme-catalysed (fungal and animal) methods.



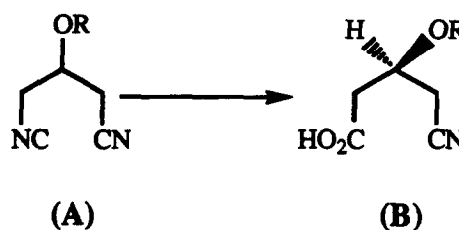
Microbial Hydrolysis of Glutatonitrile Derivatives with *Brevibacterium* Sp R312

A. Kerridge, J. S. Parratt, S. M. Roberts, F. Thiel, N. J. Turner and A. J. Willetts

^aDepartment of Biological Sciences, University of Exeter, Exeter, Devon EX4 4QG, U.K. ^bDepartment of Chemistry, University of Exeter, Exeter, Devon EX4 4QD, U.K. ^cZentrum für Selektive Organische Synthese, Rudower Chaussee 5, D-12484-Berlin-Adlershof, Germany

BioMed. Chem. 1994, 2, 447

The mechanism of conversion of dinitriles (A) into cyanoacids (B) using *Brevibacterium* R312 is discussed.



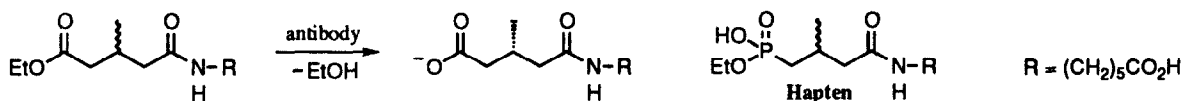
**CHARACTERIZATION OF A CATALYTIC ANTIBODY FOR
STERESELECTIVE ESTER HYDROLYSIS
-A CATALYTIC RESIDUE AND MODE OF PRODUCT INHIBITION-**

BioMed. Chem. **1994**, *2*, 457

Takuji Nakatani¹, Jun Hiratake¹, Akihiro Shinzaki², Ritsuko Umeshita¹, Tadao Suzuki², Hiroshi Nakajima², and Junichi Oda^{1,*}

¹Institute for Chemical Research, Kyoto University ²Biochemistry Department, R&D Center, Unitika, Ltd. Uji, Kyoto 611, Japan

An esterolytic antibody was generated and characterized in terms of kinetic behavior, stereoselectivity, catalytic residue, and product inhibition. A keyrole of a catalytic Arg in product inhibition is noted.



**Effect of Conformation of the Substrate on Enzymatic
Decarboxylation of α -Arylmalonic Acid**

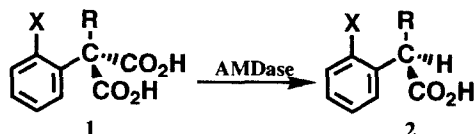
BioMed. Chem. **1994**, *2*, 469

Kenji MIYAMOTO, Hiromichi OHTA,

Department of Chemistry, Keio University, 3-14-1, Hiyoshi, Kohokuku, Yokohama 223, Japan

Yoshihiro OSAMURA,

Department of Chemistry, Rikkyo University, 3-34-1, Nishiikebukuro, Toshimaku Tokyo 171, Japan



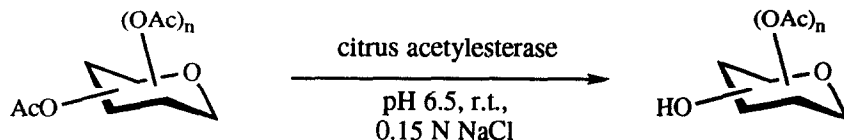
Syn-periplanar conformation of 1 is proposed to be essential to smooth enzymatic decarboxylation of 1 to afford chiral monocarboxylic acid 2.

**ACETYLESTERASE FROM ORANGE PEEL AS
BIOCATALYST FOR THE CHEMO- AND REGIOSELECTIVE
DEPROTECTION OF CARBOHYDRATES**

BioMed. Chem. **1994**, *2*, 477

Herbert Waldmann* and Axel Heuser

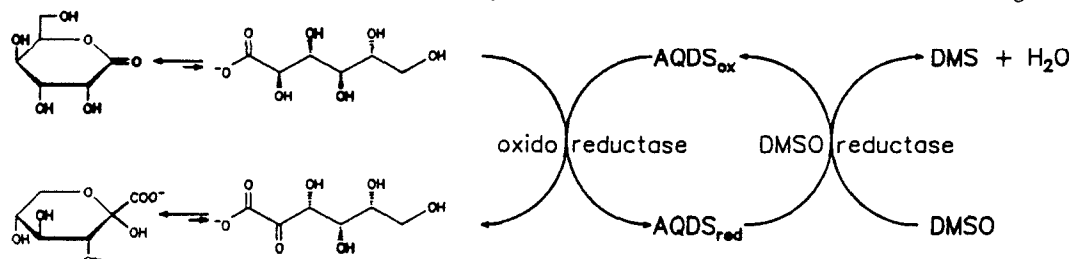
Universität Karlsruhe, Institut f. Organ. Chemie, Richard-Willstätter-Allee 2, D-76128 Karlsruhe



***P. Mirabilis* Dehydrogenates Aldonates and Aldarates with an (R)-
Configured α -Carbon Atom to the Corresponding 2-Oxocarboxylates**

BioMed. Chem. **1994**, *2*, 483

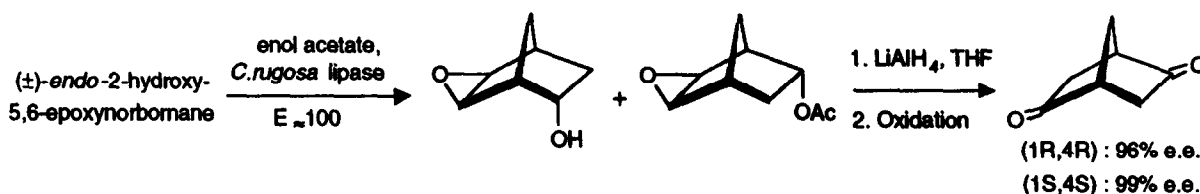
C. Schinschel and H. Simon, *Institute of Organic Chemistry, Technische Universität München, D-85747 Garching*



CHEMOENZYMATIC ACCESS TO ENANTIOMERIC BICYCLO[2.2.1]HEPTAN-2,5-DIONES

BioMed. Chem. 1994, 2, 493

Alexandra Weissfloch and Robert Azerad*, Laboratoire de Chimie et Biochimie Pharmacologiques et Toxicologiques, URA 400 CNRS, Université René Descartes, 45 rue des Saints-Pères, 75270-Paris Cedex 06, France.



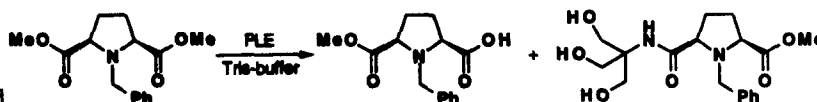
Enhanced Stereoselectivity in Pig Liver Esterase Catalysed Diester Hydrolysis. The Role of a Competitive Nucleophile

BioMed. Chem. 1994, 2, 501

Anders Mattson,^a John Boutelje,^b Ingeborg Csöregi,^c Mats Hjalmarsson,^a Ulla Jacobsson,^a Mariane Lindbäck,^a Torbjörn Norin,^a Peter Szmulik^a and Karl Hult^{b*}

^aDepartment of Chemistry, Organic Chemistry, Royal Institute of Technology, S-100 44 Stockholm, Sweden ^bDepartment of Biochemistry and Biotechnology, Royal Institute of Technology, S-100 44 Stockholm, Sweden ^cDepartment of Structural Chemistry, Arrhenius Laboratory, University of Stockholm, S-106 91 Stockholm, Sweden

The increased enantioselectivity of pig liver esterase catalysed hydrolysis of *cis*-N-benzyl-2,5-bis(methoxycarbonyl)-pyrrolidine, performed in media buffered with tris-(hydroxymethyl)aminomethane (Tris), was explained by the preference of Tris to react faster with one of the diastereomeric acyl enzymes over the other. An acid-amide product was isolated, and NMR-studies revealed the presence of an ester-amide intermediate.



DO CARBOHYDRATES PLAY A ROLE IN THE LIGNIN PEROXIDASE CYCLE?

BioMed. Chem. 1994, 2, 509

redox catalysis in the endergonic region of the driving force

Hans E. Schoemaker^{1,*}, Taina K. Lundell², René Floris³,

Tuomo Glumoff⁴, Kaspar H. Winterhalter⁵ and Klaus Piontek⁵

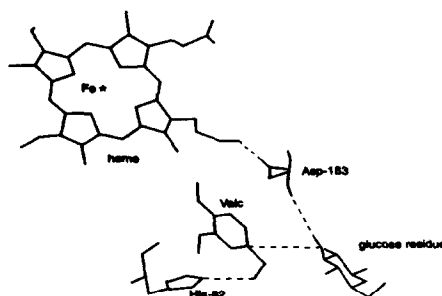
¹DSM Research, Bio-organic chemistry section, P.O. Box 18, 6160

MD Geleen, The Netherlands; ²University of Helsinki, Finland;

³University of Amsterdam, The Netherlands;

⁴University of Turku, Finland; ⁵ETH, Zürich, Switzerland;

Abstract: The redox cycle of lignin peroxidase is described in terms of Marcus theory of electron transfer. The oxidation of veratryl alcohol occurs in the endergonic region of the driving force, still the reduction of LiP Compound I is irreversible. A reversible reaction of the incipient veratryl alcohol radical cation with a glucose residue located at the entrance of the active site channel is postulated.



TWISTED α -KETO AMIDES AS TRANSITION-STATE ANALOGUES FOR ACYL-TRANSFER REACTIONS: SYNTHESIS OF THE IMMUNOCONJUGATES

BioMed. Chem. 1994, 2, 521

Jari Yli-Kauhaluoma[#] and Kim Janda^{*}

[#]On leave from Technical Research Centre of Finland,

Chemical Laboratory, Biologinkuja 7, FIN-02150 Espoo, Finland

^{*}The Scripps Research Institute, Departments of Molecular Biology and Chemistry, 10666 North Torrey Pines Road, La Jolla, CA 92037

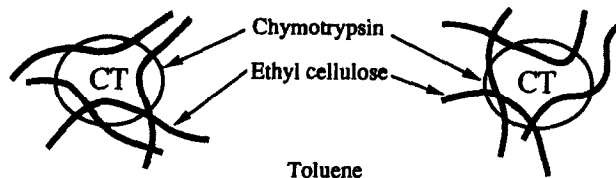
Two α -keto amides 1 and 2 were employed as antigens to elicit antibodies for acyl-transfer reactions. The rationale for their design, synthesis, and immunization protocol will be discussed.



SYNTHETIC POLYMERS AS SOLUBILIZING VEHICLES FOR ENZYMES IN WATER-POOR MEDIA.

Patrick Adlercreutz, Bo Mattiasson, Marina Otamiri
Department of Biotechnology, Chemical Center, Lund University, Lund, Sweden

Non-covalent enzyme-polymer complexes were formed by freeze-drying aqueous solutions of enzymes and polymers. Complex formation made it possible to solubilize the enzymes in organic solvents and thereby act as catalysts for the conversion of hydrophobic compounds.



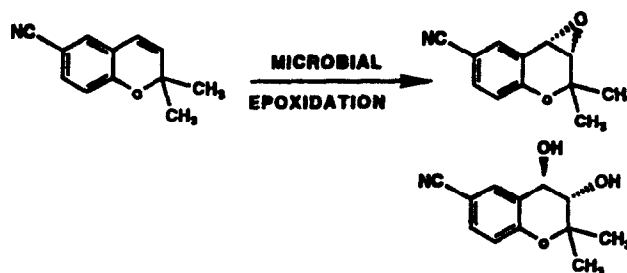
BioMed. Chem. 1994, 2, 529

Stereoselective Epoxidation of 2,2-Dimethyl-2H-1-benzopyran-6-carbonitrile

Ramesh Patel*, Amit Banerjee, Brian Davis, Jeffrey Howell, Clyde McNamee, David Brzozowski, Jeffrey North, David Kronenthal and Laszlo Szarka

Department of Microbial Technology, Bristol-Myers Squibb Pharmaceutical Research Institute, P.O. Box 191, New Brunswick, NJ 08903

Mortierella ramanniana SC 13840 and *Corynebacterium* sp. SC 13876 were used to prepare (3S,4S-cis)-3,4-Dihydro-2,2-dimethyl-2H oxireno [c] [1] benzopyran-6-carbonitrile and (3S,4R)-3,4-dihydro-3,4-dihydroxy-2,2-dimethyl-2H-1-benzopyran-6-carbonitrile

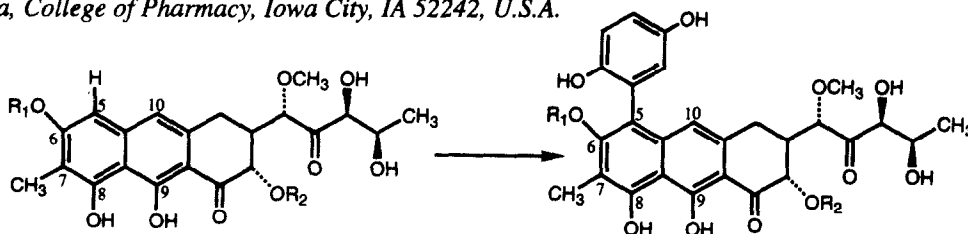


BioMed. Chem. 1994, 2, 535

Oxidative Coupling of Mithramycin and Hydroquinone Catalyzed by Copper Oxidases and Benzoquinone. Implications for the Mechanism of Action of Aureolic Acid Antibiotics.

I. O. Anyanwutaku, R. J. Petroski and J. P. N. Rosazza

Division of Medicinal and Natural Products Chemistry, and Center for Biocatalysis and Bioprocessing, University of Iowa, College of Pharmacy, Iowa City, IA 52242, U.S.A.



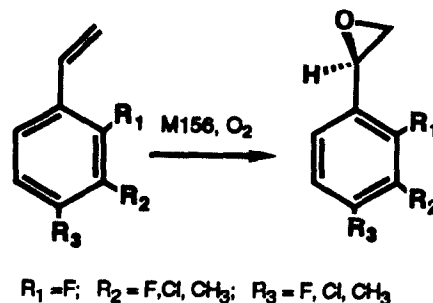
BioMed. Chem. 1994, 2, 543

Epoxidation of Styrene and Substituted Styrenes by Whole Cells of *Mycobacterium* sp M156

Stuart R. Rigby,¹ Collette S. Matthews² and David J. Leak²
¹ZENECA Bio Products, P.O. Box 2, Belasis Avenue, Billingham, Cleveland, TS23 1YN, U.K.

²Centre for Biotechnology, Department of Biochemistry, Imperial College of Science Technology and Medicine, London SW7 2AZ, U.K.

Propene grown *Mycobacterium* sp M156 catalyses the stereospecific conversion of styrene and ring mono-substituted styrenes to their epoxides in high enantiomeric excess, at rates comparable to that of the natural substrate, propene. Substituents closer to the site of epoxidation reduce the rate of reaction.



BioMed. Chem. 1994, 2, 553